

Introduction

The 8-channel Octet RED96 system from ForteBio uses biolayer interferometry (BLI) to characterize biomolecular interactions in real-time, giving insight into desirable kinetic parameters such as k_{on} , k_{off} and k_d for a given interaction.

Additionally, the Octet can be used for applications ranging from protein quantitation, lead identification/optimization, antibody characterization and cell line development.

Features

Dip and Read Technology

- The biosensor tip is coated with a compatible matrix that minimizes non-specific binding while providing a uniform and non-denaturing surface for biomolecules.

Wide range of samples

- The Octet is suitable for use with purified samples as well as unpurified samples (including whole cells) with multiple complex matrices.

Label free study of proteins

- Samples do not need to be chemically labelled for detection, allowing for the study of samples in their native context, giving data that is more representative of conditions *in situ*.

Fast, simplified workflows

- Workflows can be simplified to five steps that are heavily automated, allowing for high throughput analysis of 96 well plates in under 30 minutes.

Non-destructive investigation of samples

- Given the non-invasive nature of biolayer interferometry, samples can be recovered and used for further experiments, cutting down on the amount of protein needed.

User friendly software

- Software makes setting up runs simple, allows for modification of protocols mid run and makes data processing straightforward.



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Octet RED96 Biolayer Interferometry

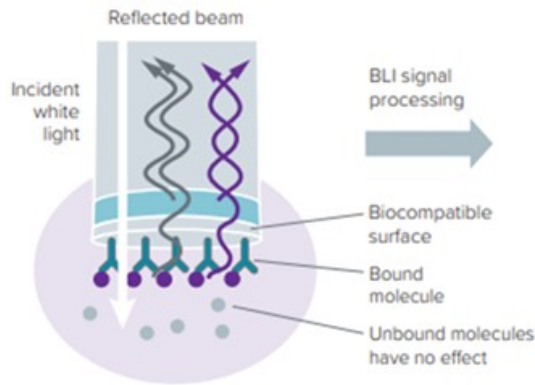
Biomolecule Quantitation and Molecular Interaction Kinetics

- Determine k_{on} , k_{off} , k_d for desired biomolecular interactions in real time.
- Detection and quantitation of analytes ranging in size from hundreds of nm (bacteria/viruses) to low nm (antibodies/peptides).
- High throughput analysis of samples with 8 parallel channels.
- Wide variety of biosensor tips permitting multiple ways to immobilize target sample.

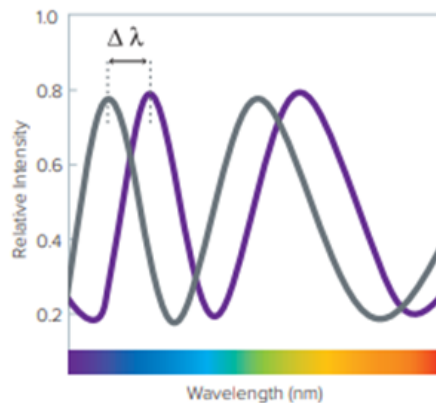
Octet RED96 Biolayer Interferometry

Theoretical Background

Biolayer Interferometry is an optical technique that detects changes in interference patterns from light waves. A biomolecule of interest is immobilized onto the tip of a biosensor and light interference at the tip of the biosensor is measured. The incident light reflects through two layers: a biocompatible layer and an internal reference layer.



Biological interactions will alter the thickness of the biolayer at the biosensor tip, thus altering the way the light reflects, resulting in a spectral shift that can be detected and monitored via a sensogram. The resulting spectral shift is then measured and can provide real time insight into the kinetics of the molecular interaction.

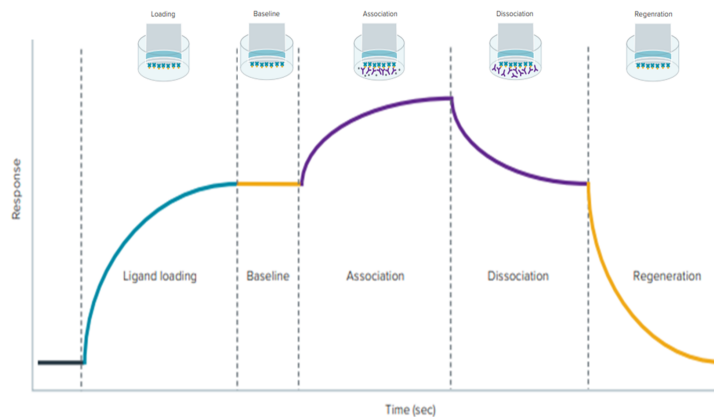


Example workflow

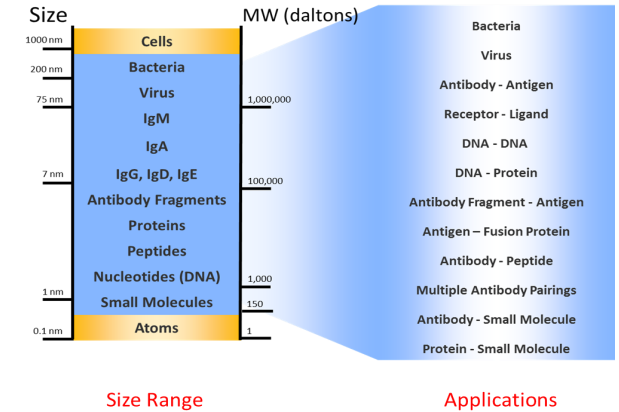
Below is a summary of a typical Octet experiment.

- 1) Loading is the immobilization of one binding partner (ligand) using the appropriate immobilization strategy.
- 2) Baseline is the washing away of any loosely bound ligand to equilibrate the biosensor tip.
- 3) Analyte association introduces the ligand's binding partner in order to obtain association kinetics.
- 4) Dissociation of the analyte from the formed complex obtains dissociation kinetics.
- 5) Regeneration of the biosensor removes bound ligand to allow for further experiments if applicable (such as when using Ni-NTA tips).

Idealized data for what one should expect to see for a typical kinetics experiment is shown below. Plotted is the response (the magnitude of the wavelength shift as a result of binding to the biosensor) as a function of time. The resulting data can then be fit to appropriate models to get the desired information.



Applications and size ranges compatible with Octet



Immobilization strategies

The table outlines a selection of strategies (others are available) for immobilizing samples based on sample features.

Sensor	Description	Mechanism of action
AHC	Anti-hIgG FC	Binding to human Fc-containing proteins and target analytes.
AR2G	Amine Reactive 2G	Binding to lysine residue of sample
NTA	Ni-NTA	Nickel binding to His tags
SA	Streptavidin	Binding to biotin tagged sample
SSA	Super Streptavidin	Binding of biotin tagged protein for small molecule screening

Technical properties of the Octet

Sample volume	180-220 μ L
Analysis temperature	15-40°C
Association rate constant (k_a)	10^2 to 10^5
Dissociation rate constant (k_d)	10^{-5} to 10^{-2}
Affinity constant (K_D)	0.1 mM – 0.1 nM
Molecular weight detection	>150 Da